

J. Environ. Treat. Tech. ISSN: 2309-1185

Journal web link: https://dormaj.org/index.php/jett https://doi.org/10.47277/JETT/10(1)109



# **Biochemical Mechanisms of True Mangrove Plants** under Salinity Stress - A Review

# A. Arunprasath\* and V. Sankara Vel

PG and Research Department of Botany, PSG College of Arts & Science, Coimbatore, Tamil Nadu, India - 641 014

Received: 04/01/2021 Accepted: 10/08/2021 Published: 20/03/2022

#### **Abstract**

In the current evaluation, mangrove plants grow in salinity; have an exceptional system of adaptation, via the tropical and subtropical estuaries. The current estimate of the world's mangrove forests is not exactly half the contrast with its origin; due to the various anthropogenic activities and climatic changes, this is expected. The accomplishment of the mangroves to grow up and thrive under brutal conditions is through their outstanding morphological, anatomical, physiological, and biochemical highlights. According to antioxidative enzymes and improvements in chloroplast structure and function, the effects of salinity on mangroves have been focused. Biochemical studies have shown that plants under salinity accumulate different metabolites called compatible solutes because they do not interfere with the metabolism of the plant and the accumulation of these solutes help as osmoprotectant in plants. It was found in different plant species that salinity stress caused the collection of soluble sugars, proline, and proteins in leaf growth to be generally restricted under salinity. Salinity causes several venomous effects, such as photosynthetic rate obstruction, chlorophyll material, and damage to the penetrability of the plasma layer, and other metabolic influences. The determination of different biochemical boundaries shows the salinity tension, which is ubiquitously seen in India in the seedling of various true mangroves, notwithstanding photosynthetic pigment stabilization. Due to the high salinity content and minerals within these plants, mangrove plants are more and more susceptible to oxidative stress and damage. This study was completed to investigate the impact of salinity on different biochemical parameters, particularly NaCl.

Keywords: Mangroves, Biochemical constituents, Salinity, Protein, Osmolytes

### 1 Introduction

A combination of the Portuguese word "Mangue" and the English word "grove" is known to be the word "Mangrove". Mangrove plants normally grow in a particular salinity climate, so they have a special adaptation system and are spread over the tropical and subtropical estuaries [1-3]. The unique regions are called the 'mangrove zone' where these plants occur. Where these plants occur, the special regions are called the 'mangrove zone'. Mangroves are usually spread over a total of 150,000 sq. km in more than 123 nations. A total of 70 all-inclusive species of true mangroves are recorded and 39 types of true mangroves are recorded in India and 86 types of associated mangroves. It is estimated that the entire mangrove area along the Indian coastline is approximately 700,000 ha, and India accounts for about 45.8 percent of the total distribution of mangroves in South Asia. Mangroves are generally spread over a total of 150,000 sq. km in more than 123 nations. A total of 70 allinclusive species of true mangroves are recorded and 39 types of true mangroves are recorded in India and 86 types of associated mangroves. It is estimated that the entire mangrove region along the Indian coastline is approximately 700,000 ha, and India accounts for about 45.8 percent of the total distribution of mangroves in South Asia. The mangroves along India's east coast are progressively (80%) than the western coast (20%) because the territory on the east coast has a steady slope as fields as opposed to the lofty slope along the west coast. The Sundarbans in West Bengal cover half of the total mangrove territories in India. The latest estimate of the world's

mangrove plantations is not quite 50% of what they used to be [4, 5]. The primary purpose behind this situation is to increase segment pressure and climate change factors, such as ocean level rise, elevated salinity, highly distractive typhoon visits, tsunami, and so on, particularly because industrialization, is considered to be the best risk to the economic and biological usefulness of plant networks all around [6, 7].

True mangroves are "tropical trees restricted to intertidal and adjacent communities" [8], growing in saline, usually in anoxic soils, under climatic conditions defined by a blend of high temperature and irradiance [9]. true mangroves possess any or the overwhelming majority of the accompanying highlights: (i) occurring only in mangrove environment and not extending into terrestrial communities; (ii) morphological specialization (aerial roots, vivipary); (iii) physiological mechanism for salt exclusion and/or salt excretion; (iv) taxonomic isolation from terrestrial relatives. Tomlinson's measures gave an extremely 'clear' norm to organize true mangroves and mangrove associates and it has been accepted generally [10-14] and more taxa have been classified clearly. It's unique morphological, anatomical, physiological, and biochemical highlights are the achievement of the mangroves to develop and thrive under harsh conditions [15, 16]. It is recommended that the viviparous condition, commonly found in the Rhizophoraceae family, has extraordinary flexible adaptive significance to survive in the intertidal regions by maintaining a strategic distance at the germination stage from high salinity.

Corresponding author: A. Arunprasath, PG and Research Department of Botany, PSG College of Arts & Science, Coimbatore, Tamil Nadu, India – 641 014, E-mail: arunprasath194@gmail.com

The effects of salinity on mangroves were concentrated in terms of changes in chloroplastic structure and capacity according to anti-oxidative enzymes [17, 18]. Salinity causes several venomous effects, including photosynthetic rate inhibition, chlorophyll content, plasma membrane permeability damage, and other metabolic disorders [19] Biochemical studies have shown that plants under salt pressure collect different metabolites called compatible solutes because they do not interfere with the metabolism of plants and the accumulation of these solutes contributes to the maintenance and osmoprotectant of turgor in plants. In different plant species, it has been found that salinity stress has generally inhibited the accumulation of solvent sugars, proline, and proteins in leaf development under salinity [20]. The significant free amino acids acquired in the leaves of the mangroves are aspartic corrosive, alanine, proline, tryptophan, tyrosine, and phenylalanine [21]. Amino acid existed under salinity stress in the plant tissues but was additionally submerged in higher plants with water stress [22]. The dynamic decrease in protein and rise in the total free amino acids under the salinity of NaCl was either due to the change of protein into amino acids or the limitation of the consolidation of amino acids into protein [23]. A gathering of free amino acids was also reported a lot of free amino acids, yet have a decrease in protein content during an exposing time of high-saline introduction [24]. In their leaves, mangroves store high concentrations of proline and glycine betaine [25]. In halophytes, proline accumulation is an overall wonder. It is noteworthy that many stresses, including salt stress, increase the proline content in the leaves of many plants [26, 27]. The most commonly known perfect solvent that ensures the photosynthetic apparatus of plants is glycine betaine [28]. Mangroves are salt-excreting plants that are more susceptible due to high salt content to oxidative stress and damage. Therefore, self-defense in the form of high antioxidants in these plants against this oxidative stress emerged [29]. It is generally accepted that by increasing the activity of cancer prevention agent chemicals, larger plants oppose antioxidants. There were also a few reports on the relationship between salinity and cancer prevention agent compounds in mangrove plants [30]. Owing to the proximity of large measurements of phenolic blends, the cancer prevention agent properties of these mangrove plants are attributed. Terpenoids, phytoalexins, coumarin subordinates, tannin and its subsidiaries, flavonoids, and their subsidiaries [31, 32]. In plants including phenols, flavonoids, and so on, some optional metabolites have important work against pressure conditions [33]. Salt stress (50 and 100 mM NaCl) practically produces grain flavonoid content [34].

In a biological context, ROS, explicitly hydrogen peroxide (H2O2), goes about as signaling molecules; however, in outrageous conditions, their levels essentially increment and damage cell functions, in the end prompting mortality if the horrible condition perseveres [35]. Flavonoids reduced the amount of oxygen from UV radiation and preserved mangroves [36]. As critical cell reinforcements or free radical scavengers and antimicrobials, flavonoids are a major accumulation of optional exacerbates [37]. The reactive oxygen species that cause layer dysfunction and cell lethality are side effects of hyperosmotic and ionic stresses [38]. ROS, explicitly hydrogen peroxide (H2O2), goes around as signaling molecules in a biological context; however, their levels effectively increase and destroy cell functions in outrageous circumstances, inevitably prompting mortality if the awful condition continues [39]. H<sub>2</sub>O<sub>2</sub> acts as an indicator molecule for controlling the typical salinity response [40]. Then again, plants have a wonderful ability to process H2O2 as distinct creatures, and cells treated with a focus as high as 10 mM can use the

compound entirely in under 10 minutes [41]. The increased development of high AOS is associated with increasing evidence of current membrane injury under salt stress. Since lipid peroxidation is the easiest symptom of oxidative harm, it is sometimes used as an oxidative damage indicator [42]. Lipid peroxidation has been measured as MDA formation as an indicator of membrane integrity. A significant number of recent studies have focused on the morphological, physiological, and biochemical characteristics of the salinity or salt pressure response of grown plants, including problematic instruments [43].

In the current examination, a challenge has been completed towards the study in the effect of salinity on different biochemical parameters were reviewed on some of the Indian true mangroves such as Aegiceras corniculatum (L.) Blanco, Avicennia alba Blume, Avicennia marina (Forsk.) Veierh, Avicennia officinalis L., Bruguiera cylindrica (L.) Blume, Bruguiera gymnorrhiza (L.) Lamk, Bruguiera parviflora Wt.& Am., Ceriops decandra (Griff), Ceriops roxburghiana Arn., Ceriops tagal (Per.)Rob, Excoecaria agallocha L., Heritiera fomes Buch-Ham, Heritiera littoralis Dry and ex Ait., Kandelia candel (L) Druce, Kandelia obovata (S., L.) Yong, Phoenix paludosa Roxb, Rhizophora apiculata Blume, Rhizophora mucronata Lamk, Rhizophora stylosa Griff, Sonneratia alba J. Smith, Sonneratia apetala Buch-Ham, Sonneratia caseolaris (L.) Engler and Xylocarpus granatum Koenig [44-46].

#### 2 Literature review

#### 2.1 Photosynthetic pigments under Salinity Stress

At high salt treatment concentrations at 30 d Chl a: b ratio, Aegiceras corniculatum yielded more. It, therefore, gives the impression that even though the overall Chl content decreased at high salt concentration, the high salt concentration did not affect Chl a: b proportion [47]. While the salinity of Avicennia officinalis leaves advanced up to 0.75 percent in chlorophyll synthesis. However, a difference in the concentration of sodium chloride in the photosynthesis of the net leaves was found in the leaves A. officinalis. There was a decrease in chlorophyll content at higher sodium chloride concentrations [48]. The absolute chlorophyll content in leaves showed a decreasing example in Bruguiera cylindrica, after increasing salinity concentrations, but there was no significant shift in the complete chlorophyll content of salt-treated plants [49]. The overall Chl and carotenoid content of Bruguiera parviflora changed fundamentally when different NaCI concentrations were added in plants treated with a high salt concentration, with a higher proportion. In this way, even though the total Chl substance increased at low levels and decreased at higher concentrations, these high salt concentrations do not affect this ratio [50]. Ceriops roxburghiana has shown that NaCl revives the synthesis of chlorophyll to an ideal concentration of 300 mM. There was a drop in the content of chlorophyll and carotenoids at higher concentrations [51]. In the dry season, photosynthesis rates were essentially lower than in the wet season in Ceriops tagal and Rhizophora mucronata [52]. In comparison to control plants, the effects of NaCl on Kandelia candel pigments after momentary 250mM NaCl treatment were higher for chlorophyll and carotenoid substances in treated plants. Moreover, in plants treated with 250 mM NaCl, photosynthetic pigments aggregated during long-term NaCl treatment reached the most important amount and the lowest level in plants measured with 500 mM NaCl [53]. The chlorophyll content in non-saline conditions was higher in Bruguiera gymnorrhiza, Excoecaria agallocha, and Heritiera fomes. In the previous two, the rise in absolute chlorophyll was estimated to be enormous as salinity decreased in the soil when

in *Heritiera*. Interestingly, the complete chlorophyll was initiated by *Phoenix paludosa* and *Xylocarpus granatum* with increased soil salinity. Nevertheless, in plants grown in nonsaline soil, the ratio of Chlorophyll a and b is higher. In any event, there was not much effect on the chlorophyll material, but rather a decrease in the amount of the substance. High salinity has repressed photosynthesis. Due to the high concentration of salinity in *Rhizophora stylosa*, *Sonneratia apetala*, and *Sonneratia caseolaris*, the net photosynthesis rate is decreased. The decreasing trend was evident at the point where salinity increased highly [54]. The decrease in the Chl content at high NaCI concentrations may be due to the interruption of certain chloroplasts or changes in the proportion of pigment-protein complexes in the lipid-protein, just as given increased chlorophyll action [55].

# 2.2 Sugars and Starch content under Salinity Stress

At higher NaCl concentrations (250 mM), the total sugar content of the corniculatum decreased by 1.9-fold during 30 d of treatment. At 250mM NaCl, the starch then again increased by 2.7-folds towards the end of 30 d. Besides, the degree of adjustments in the control and 250 mM NaCl treated plants was measured to reduce the sugar proportion. Even though the ratio of no reduction to sugar reduction increased significantly through salt treatment in A. Corniculatum, to find out their shift to salt assurance in this species with the screening of sugar alcohol levels would be of some importance [47]. The content of soluble sugars in B. cylindrica leaves and root tissues have decreased when confronted with increased NaCl classification. In the leaf tissue of plants exposed to 500 mM NaCl fixation, the complete solvent sugar increased. There was a continuous decrease in total dissolvable sugar in 400 mM, 500 mM, and 600 mM NaCl concentration separately over the controlled plants in both leaf and root tissues over 20 days of therapy [48]. Both the starch and sugar compound B. parviflora only increased marginally after 45 d. Treatment with 400 mM NaCI caused a 2.5-overlap increase in the total sugar content during this equivalent period. Starch content decreased by 40 to 45 percent with 400 mM NaCI on Day 45, on the other hand. The total sugar content increased to a low level at 100 mM and the level of starch decreased, but with 200 mM of NaCI, the changes in sugar and starch levels were similar to those registered at 400 mM. This suggests prompt changes in sugar/starch transformation at a low concentration of NaCl. Even though because of salt treatment in B. parviflora, this proportion did not appear to change much. To learn their transformation towards salt protection in this species, the level of sugar alcohols should be examined later [50]. [51] Found adjustments in the C. roxburghiana. The total sugar content in leaves (6.50 and 7.50 mg/g fr. wt.) and roots (2.96 and 5.18 mg/g fr. wt.) was gradually reduced to an ideal amount (300 mM) in leaves and roots on the 30th and 60th days respectively, in response to different NaCl concentrations. The overall sugar content increased with a higher salt concentration. Information on the starchy substance of leaves (5.12 and 6.15 mg/g fr. wt.) and roots (4.10 and 5.95 mg/g fr. wt.) at different salinity levels increased separately in leaves and roots up to 300 mM with increased salinity on 30th and 60th days. After long-term culture with 250 and 500 mM NaCl, the K. candle soluble sugar substance persisted at more substantial levels [53] The content of the starch was found higher in S. alba as the NaCl concentration expands, [56]. There was a gradual decline in starch at higher concentrations. The absolute soluble sugar content of S. caseolaris and S. apetala leaf is elevated in high salinity by a greater amount than in low salinity. In the leaves, the total solvent sugar content of R. stylosa has steadily increased (Yan L, and Guizhu C, 2007). Starch and sugar levels

are impacted by stress, like other cell constituents [57]. [58] Announced that salinity lowers the salinity of solvent and hydrolyzable sugars. In *Rhizophora stylosa, Sonneratia apetala*, and *Sonneratia caseolaris*, the net photosynthesis rate is reduced due to high salinity. The reduction tendency was self-evident at the stage when salinity increased high [54]. Due to the disruption of some chloroplasts or changes in the lipid-protein ratio of pigment-protein buildings, the decrease in Chl content at high NaCI fixations may be due to increased chlorophyll action [55].

#### 2.3 Amino Acid under Salinity Stress

The free amino corrosive substance of A. corniculatum was diminished altogether during treatment with 250 mM NaCl [47]. The level of an event of free amino acids were decreased in the plants developed in non-saline condition, for example, 33.47% in B. gymnorhiza, 9.21% in E. agallocha, 55.4% in H. fomes 33.22% in P. paludosa and 24.1% in X. granatum. In both the leaf and root tissues of B. cylindrica, the most noteworthy amino acid content was reported, rise in NaCl concentration, amino acid increase [49]. In the control tests for B. parviflora, the free amino acids did not shift. However, total amino acids increased only slightly at the 100-mM treatment stage, while their expansion was consistent at 200 and 400 mM [50]. Changes were observed in the absolute free amino acid at different degrees of salinity of NaCl. The amino corrosive compound in the leaf and the base of C. roxburghiana. With increased NaCl concentrations, decreased by up to 600 mM [51]. The amino acid content R. apiculata decreased fixation by up to 400 mM and increased at higher salinity levels. When the amino acid content showed a decrease in the ideal salinity of 400 mM, it reported a steady increase at higher concentrations [59]. Besides, several free amino acids have been detailed in a series of free amino acids, but during an all-encompassing period of high-saline presentation, they have a decrease in protein content [24].

#### 2.4 Protein content under Salinity Stress

The protein substance of A. corniculatum indicated a statistically significant, yet exceptionally small, decline upon salt treatment. Though SDS-PAGE examination demonstrated no significant changes in protein profiles of control and salttreated samples at these shows there are no progressions or exceptionally mild changes in the protein content [47]. In R. apiculata protein level increased up to the ideal degree of NaCl salinity and reduced at higher fixation [59]. In B. parviflora indicated decreases in protein content. A salt induces a decrease in solvent protein content proceeded for the initial 14 d of treatment, and afterward, the level stayed stable. Complete protein content gradually diminished with increasing concentrations of NaCl. This diminishing may have come about because of an adverse effect of NaCI on protein synthesis or proteolysis [50]. A few proteases are known to be initiated by salt stress [60]. Although protein content can decrease at higher salt fixations, [61] found that the degree of 33-kDa manganese balancing out protein increased in B. gymnorrhiza. At the point when the impact of NaCl on the protein content in leaves and base of C. roxburghiana are increased with increasing salinity up to 300 mM with the most extreme increment on the 30th and 60th day. The leaf had more protein than the root. Here, the expansion in the concentration of NaCl increased in the protein content [51]. It was established that the increase is differed each and profoundly found in the E. agallocha and P. paludosa. In any case, H. fomes and X. granatum indicated a minimal measure of protein degradation happen in a salt environment than the others, most likely prompting lesser capacity to synthesis good amino acids in the

salty natural surroundings. In *K. candel* the absolute protein content didn't change altogether in at first treated plants, however, in the (500 mM) higher centralization of NaCl where it is reduced to 14% [53]. Protein content in *A. marina* diminished upon introduction to 500mM NaCl for a brief time of 7 days Absolute dissolvable leaf protein content likewise diminished 3.4 overlaps following 7 days of salt treatment when contrasted with control [62]. Stress instigated proteins amassed in the cell which may be synthesized because of the salt or maybe available constitutively at a low level [63]. The peripheral change in protein substance and protein profile recommends that NaCl presentation influences the exceptionally diminished protein substance of the leaves [58, 64]

#### 2.5 Osmolytes under Salinity Stress

The proline substance of A. corniculatum leaves diminished by 75% in 250 mM NaCl treated plants when contrasted with control however there is no change in controlled samples during the investigation [47].In A. marina proline content was increased in the two seedlings and saplings under salt stress yet its relative commitment to the total osmolality was most minimal [65]. Proline content in leaf and root of B. cylindrica increased after increasing the NaCl concentration. Proline content was increased to the extent of 39%, 58%, and 124% with 400 mM, 500 mM, and 600 mM NaCl treatment in leaf tissue, and at the root, the expansion was about 12%, 39%, and 93% in 20 days after the control plants [49]. Free proline content was identified in B. gymnorrhiza, P. paludosa, and X. granatum, whereas it is absent in H. fomes and E. agallocha. In P. paludosa, proline content was generally high, while a lower concentration of proline was assessed in B. gymnorrhiza and X. granatum during Salinity. Proline amalgamation in light of salt pressure increased to the most extreme level the plants required 14 d to acclimate to the adjustment in the outside NaCI content; after which, the cell content level stayed consistent. Hence, proline accumulation is a significant salt-stress versatile component in B. parviflora [50]. In C. roxburghiana, proline accumulation increased fundamentally under the different increasing concentrations of NaCl [51]. In K. Candel proline levels increasing in both the 250 and 500 mM NaCl concentration long-term culture when contrasted with the control plants [53]. The collection of proline as either a compatible osmolyte in NaCI stress or for an ion ratio imbalance. Accumulation likewise happens when the action of proline dehydrogenates, a catabolic enzyme, is repressed [66].

#### 2.6 Phenolic and Flavonoid contents under Salinity stress

NaCl treatment increased the polyphenol activity of A. corniculatum leaves, which was 10.3% higher than the control [67]. The measure of total phenolics in evident mangroves A. corniculatum, A. Alba, B. gymnorrhiza, C. decandra, R. mucronata, and S. apetala were ranging from 4.40 to 94.41 mg GAE/g dry material. Among leaves, the most elevation was found in S. apetala (47.52 GAE) and the most minimal in C. decandra (5.14 GAE). The stem bark of mangroves contained a significant measure of phenolics. The highest was found in C. decandra (94.41 GAE) and the least in the stem bark of A. alba (4.40 GAE). Among root materials, C. decandra was found to contain the most elevated phenolic segment (73.60 GAE) and the least was found in A. alba (4.79 GAE). It was seen that three types of mangroves in the Rhizophoraceae family, phenolic substances were incredibly high in its stem bark than its root/leaves under salinity conditions [31]. The complete phenolic content was seen as increasing after increasing NaCl

fixations in both leaf and root tissues of B. cylindrical [49]. The complete phenolic substance of the two plants methanol separate was extrapolated from the calibration curve. They were  $36.625\pm0.551$  and  $58.917\pm0.601$  mg Gallic acid equivalents/g dry plant material in H. littoralis and B. gymnorrhiza, individually under salinity. Absolute flavonoid substances were extrapolated from the direct condition of the quercetin standard curve. TFC acquired for both methanol concentrates of *H. littoralis* and *B. gymnorrhiza* are 114.52 ± 0.339 and 77.21  $\pm$  0.016mg quercetin equal/g extricates under saline conditions [68]. There was a slow increment of 1%, 7%, and 42% in leaf and 3%, 10%, and 22% in root tissue when treated with 400 mM, 500 mM and 600 mM NaCl concentrates on 20 days of treatment over the control plants [49]. The total phenolic content in E. agallocha become increasing up to 300mM NaCl and afterward decreasing yet at the same time the substance stays high in contrast with control. The most maximum value was 2.125±0.15 mg GAE/gram in leaf tissue on the 28th day of NaCl presentation. The total flavonoid content in E. agallocha becomes increments up to 200mM NaCl with the maximum value 1.4±0.1 mg QE/g leaf tissue and then diminishes but at the same time the content stays high in contrast with control at 28th day of NaCl presentation [69]. In K. candel the total concentration of phenols indicated no significant difference still 200 mM NaCl treatments, but it increased 17% less than 500 mM NaCl stress. Similarly, the total concentration of flavonoids didn't increase from 0 to 200 M NaCl treatment, however increased in 500 mM NaCl treatment [70]. Phenolic and flavonoid content increased extensively in plants at increased salinity and decreased at high salinity [71].

# 2.7 Malondialdehyde and Hydrogen Peroxide content under Salinity stress

MDA, a cytotoxic result of lipid peroxidation, has been considered as a marker of oxidative damage incited by free radicals. H<sub>2</sub>O<sub>2</sub> is one of the most significant peroxides. Compared with the control, MDA and H2O2 substance increased by 20% and 15%, respectively, in NaCl-treated A. corniculatum leaves [67]. Studies in A. marina with H2O2 stress indicated a postponed reaction at mRNA level. In any case, indicated a slow increment but didn't demonstrate constant increment in m-RNA convention. Nevertheless, increased mRNA levels were seen as an increment in salinity treatment [72]. MDA substance in NaCl-treated plants of B. gymnorrhiza stayed at the same level as those in controls during about a month of stress, even 21-22% expansion was seen toward the end of the analysis (400 mM NaCl) essentially, increasing Salinity [73]. The content of lipid peroxidation in transgenic lines and control plants of A. marina was examined through measurement of malondialdehyde level. Under unstressed conditions, no significant distinction was seen in the malondialdehyde content. Introduction to salt stress caused a double increment in malondialdehyde content in charge plants, but no significant increment was seen in transgenic lines contrasted with individual controls [74]. The MDA content of B. cylindrica was seen as increased after increasing NaCl concentrations in both leaf and root tissues. The MDA content demonstrated an expansion of 6%, 22%, and 125% in leaf, and 7%, 13%, and 33% expansion was seen in root when treated with 400 mM, 500 mM, and 600 mM NaCl individually over the control plants on 20 days of treatment [49]. In B. parviflora H<sub>2</sub>O<sub>2</sub> level in the control leaf didn't show a lot of progress during the 45 d treatment, while in 400m mol/L NaCl treated plants, the H<sub>2</sub>O<sub>2</sub> level increased with increment in NaCl focuses after the 45d treatment. In both control and salt-treated tests, the grouping of MDA remained almost at a similar level. No

change in MDA level indicated that lipid peroxidation didn't happen and hence, layer uprightness was not influenced [47]. Under salinity stress, exogenously applied H<sub>2</sub>O<sub>2</sub> (10 mM) significantly accelerated the efflux of Na+ along the root axis in the two species. Progressively articulated impacts were seen in B. gymnorrhiza contrasted and K. candel [75]. During a test in K. candel, the degrees of H2O2 in treated leaves stayed unaltered in 250 mM NaCl culture. Nonetheless, the H<sub>2</sub>O<sub>2</sub> levels accumulation in 500 mM NaCl during constant treatment. The presence of lower MDA levels at 500 mM NaCl was found. This proposes K. candel has a higher limit concerning gestures of the ROS that was prompted by salt, which might be helpful to present at high saline living habitats [53]. Utilizing NaCl treated plants, the impacts of H2O2 on roots of K. obovata, no additive effect was noted. In numerous treatment under NaCl stress, on treated roots was not diminished by H2O2. Consequently, no change was found under salinity [76]. Hydrogen peroxide rummaging limit of 5 plants were estimated. In E. agallocha, B. gymnorhiza and P. paludosa indicated an expansion in the level of a hindrance though most reduced restraint in H. fomes and X. granatum [77]. At higher H<sub>2</sub>O<sub>2</sub> concentration levels, the rate of R. stylosa was fundamentally reduced compared with plants with lower H<sub>2</sub>O<sub>2</sub> concentration levels. R. stylosa at the higher rise had moderately higher H<sub>2</sub>O<sub>2</sub> concentration levels under salinity stress [78]. It has been accounted for that salt pressure can trigger the collection of reactive oxygen species ROS, for example, hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) in mangrove plants. These cytotoxic ROS can genuinely disturb metabolic activity through oxidative damage to lipids, proteins, and nucleic acids

# 2.8 Enzymatic and Non Enzymatic antioxidants systems under Salinity stress

In this examination, there is no notable change was seen in the activity of SOD and CAT in A. corniculatum leaves among various medicines. GPX movement displayed no adjustments in leaves exposed to NaCl. Be that as it may, GPX action treated with NaCl was increased gradually in NaCl-treated seedlings. On PPO movement the NaCl-treated seedling increased the activity [67]. In B. cylindrica during enzymatic movement at higher NaCl concentration (500 mM and 600 mM) all the four proteins, for example, APX, GPX, SOD, and Catalase was upgraded altogether increasing action was found in leaf and root, separately. Though in Non-enzymatic action in Ascorbate and glutathione content demonstrated a diminishing pattern with increasing NaCl medicines both in leaf and root when contrasted with control plants [49]. The movement of CAT, APX, and POX was estimated in charge of B. parviflora and salt tests. The action of CAT and APX increased by 60% in salt-treated plants, but the action of guaiacol peroxidase was not changed more. The movement of these antioxidative proteins in control tests remains nearly the equivalent in the whole time of investigation [30]. The activity of SOD of K. candel at first in momentary culture is reduced by 42% and 38% but at 500 mM NaCl concentration, this movement increased altogether, as contrasted and the control, separately. The PRX movement in leaves during short-term treatment stayed unaltered however after long-term treatment with 500 mM NaCl action was increased [53]. In this examination antioxidant activity of both PRX and SOD were high in saline plants and the increment extended from 139-257% in PRX and 147-241% in SOD found in B. gymnorhizha, E. agallocha, and P. paludosa. In any case, H. fomes and X. granatum indicated a lower rate while compared with the other three plants under saline conditions. It was to realize that peroxidation was influenced severely by salt stress in leaves and stems of S.

apetala, S. caseolaris and R. stylosa. MDA content in leaves and stems of Sa and Sc decreased marginally in low salinity and then increased quickly, which was reverse to the activity of SOD of S. apetala and S. caseolaris no more SOD activity are created, which may be identified with the sorts of the plants or potentially to the fact that S. apetala and S. caseolaris made other peroxisome or catalase protecting membrane peroxidation from being demolished with the assistance of active oxygen free radicals [54]. The salinity forced plants with elevated levels of antioxidants have certain opposition against oxidative harmless [47].

#### 3 Conclusion

The present study shows that Mangroves are moderately salt-tolerant species. They can be easily propagated under limited or low-salinity conditions. At high salt levels plants become adapted to salt stress after two to three weeks of exposure. During this adaptive period, changes occur in the levels of photosynthetic pigments, proteins, antioxidants, and other biochemical characters. It can be concluded that under salinity stress does not have strong protection ability for plant membranes. Numerous mangrove species adapted by filtering as much as 90 percent of the salt found in seawater as it enters their underlying foundations. A few species discharge salt through glands in their leaves. However, Sodium chloride salinity stimulated its growth, biomass, photosynthetic pigments, and biochemical determinations up to the optimum concentration. Some experimental evidence also shows a higher concentration of NaCl is lethal for both salt secretor and non-secretor mangroves.

### Acknowledgments

The authors are grateful to acknowledge PG & Research Department of Botany, PSG College of Arts & Science, Coimbatore for providing necessary facilities during the study.

#### **Competing Interest**

The authors report no declarations of interest.

#### **Authors' contribution**

All others have an equal amount of contributions.

#### **Ethical issue:**

Authors are aware of, and comply with, best practice in publication ethics specifically with regard to authorship (avoidance of guest authorship), dual submission, manipulation of figures, competing interests and compliance withpolicies on research ethics. Authors adhere to publication requirements that submitted work is original and has not been published elsewhere in any language.

#### References

- Connor DJ. Growth of grey mangrove (Avicennia marina) in nutrient culture. Biotropica. 1969 Dec 1:36-40. https://doi.org/10.2307/2989759.
- Downton WJ. Growth and osmotic relations of the mangrove Avicennia marina, as influenced by salinity. Functional Plant Biology. 1982; 9(5):519-28. https://doi.org/10.1071/PP9820519.
- Alongi D. The energetics of mangrove forests. Springer Science & Business Media; 2009 Jan 18.
- 4. Spalding M, Blasco F, Field C. World mangrove atlas.
- Spiers AG. Review of international/continental wetland resources. Global review of wetland resources and priorities for wetland inventory. 1999:63-104.
- Valiela I, Bowen JL, York JK. Mangrove Forests: One of the World's Threatened Major Tropical Environments: At least 35% of the area of mangrove forests has been lost in the past two

- decades, losses that exceed those for tropical rain forests and coral reefs, two other well-known threatened environments. Bioscience. 2001 Oct 1; 51(10):807-15. https://doi.org/10.1641/0006-3568 (2001)051[0807: MFOOTW] 2.0.CO; 2.
- Duke NC, Meynecke JO, Dittmann S, Ellison AM, Anger K, Berger U, Cannicci S, Diele K, Ewel KC, Field CD, Koedam N. A world without mangroves? Science. 2007; 317(5834):41-2.
- Tomlinson PB. The Botany of Mangroves Cambridge University Press London.
- Stewart GR. and Popp M, 1987. The ecophysiology of mangroves. In: R.M.M. Crawford (Ed.) Plant Life in Aquatic and Amphibious Habitats. British Ecological Society. Blackwell Sci. Publ. Oxford: 333-345.
- Duke NC. Australia's mangroves: the authoritative guide to Australia's mangrove plants. MER; 2006.
- Kathiresan K, Bingham BL. Biology of mangroves and mangrove ecosystems. Advances in marine biology. 2001 Dec; 40:84-254.
- Lacerda LD, Conde JE, Kjerfve B, Alvarez-León R, Alarcón C, Polanía J. American mangroves. In Mangrove ecosystems 2002 (pp. 1-62). Springer, Berlin, Heidelberg. https://doi.org/10.1007/978-3-662-04713-2\_1.
- Parani M, Lakshmi M, Senthilkumar P, Ram N, Parida A. Molecular phylogeny of mangroves V. Analysis of genome relationships in mangrove species using RAPD and RFLP markers. Theoretical and Applied Genetics. 1998 Sep 1; 97(4):617-25. https://doi.org/10.1007/s001220050937.
- Saenger P. Mangrove ecology, silviculture and conservation. Springer Science & Business Media; 2013 Jun 29. https://doi.org/10.1007/978-94-015-9962-7.
- Ball MC, Farquhar GD. Photosynthetic and stomatal responses of two mangrove species, Aegiceras corniculatum and Avicennia marina, to long term salinity and humidity conditions. Plant Physiology. 1984 Jan 1; 74(1):1-6. DOI: https://doi.org/10.1104/pp.74.1.1.
- Parida AK, Jha B. Salt tolerance mechanisms in mangroves: a review. Trees. 2010 Apr 1; 24(2):199-217. DOI 10.1007/s00468-010-0417-x.
- Parida AK, Das AB, Mohanty P. Defense potentials to NaCl in a mangrove, *Bruguiera parviflora*: differential changes of isoforms of some antioxidative enzymes. Journal of plant physiology. 2004 Jan 1; 161(5):531-42. https://doi.org/10.1078/0176-1617-01084
- Parida AK, Das AB, Mittra B. Effects of NaCl stress on the structure, pigment complex composition, and photosynthetic activity of mangrove *Bruguiera parviflora* chloroplasts. Photosynthetica. 2003 Jun 1; 41(2):191.
- Karimi G, Ghorbanli M, Heidari H, Nejad RK, Assareh MH. The effects of NaCl on growth, water relations, osmolytes and ion content in *Kochia prostrata*. Biologia plantarum. 2005 Jun 1; 49(2):301-4.
- Zdunek E, Lips SH. Transport and accumulation rates of abscisic acid and aldehyde oxidase activity in *Pisum sativum* L. in response to suboptimal growth conditions. Journal of Experimental Botany. 2001 Jun 1; 52(359):1269-76.
- Nandy P, Dasgupta N, Das S. Differential expression of physiological and biochemical characters of some Indian mangroves towards salt tolerance. Physiology and Molecular Biology of Plants. 2009 Apr 1; 15(2):151-60. https://doi.org/10.1007/s12298-009-0017-7.
- Treichel S. The effect of NaCl on the concentration of proline in different halophytes. Z. Pflanzenphysiol. 1975; 76:56-68.
- Devitt DA, Stolzy LH, Labanauskas CK. Impact of potassium, sodium, and salinity on the protein-and free amino acid content of wheat grain. Plant and soil. 1987 Mar 1; 103(1):101-9. https://doi.org/10.1007/BF02370674.
- 24. Gilbert GA, Gadush MV, Wilson C, Madore MA. Amino acid accumulation in sink and source tissues of *Coleus blumei* Benth. During salinity stress. Journal of Experimental Botany. 1998 Jan 1; 49(318):107-14. https://doi.org/10.1093/jxb/49.318.107.
- Mickelbart MV, Peel G, Joly RJ, Rhodes D, Ejeta G, Goldsbrough PB. Development and characterization of near isogenic lines of sorghum segregating for glycine betaine accumulation. Plant Physiol 2003; 118:253-61. https://doi.org/10.1034/j.1399-3054.2003.00106.x.
- Lee TM, Liu CH. Correlation of decreased calcium contents with proline accumulation in the marine green macroalga *Ulva fasciata*

- exposed to elevated NaCl contents in seawater. Journal of Experimental Botany. 1999 Dec 1; 50(341):1855-62. https://doi.org/10.1093/jxb/50.341.1855.
- Hernandez S, Deleu C, Larher F. Proline accumulation by tomato leaf tissue in response to salinity. Comptes rendus de l'Academie des sciences. Serie III, Sciences de la vie. 2000 Jun; 323(6):551-7. DOI: 10.1016/s0764-4469(00)00167-0
- 28. Papageorgiou GC, Murata N. The unusually strong stabilizing effects of glycine betaine on the structure and function of the oxygen-evolving photosystem II complex. Photosynthesis Research. 1995 Jun 1; 44(3):243-52. https://doi.org/10.1007/BF00048597.
- Nirjhar D, Paramita N, Chandan S, Sauren D. Salinity impact on the precarious Mangroves: a biochemical study on some taxa from Indian sundarbans. American Journal of Plant Physiology. 2012; 7(2):53-69. DOI: 10.3923/ajpp.2012.53.69.
- Mishra S, Bandhu AD. Effect of short-term exposure to NaCl on photochemical activity and antioxidant enzymes in *Bruguiera* parviflora, a non-secretor mangrove. Acta Physiologiae Plantarum. 2004 Sep 1; 26(3):317.
- Banerjee D, Chakrabarti S, Hazra AK, Banerjee S, Ray J, Mukherjee B. Antioxidant activity and total phenolics of some mangroves in Sundarbans. African Journal of Biotechnology. 2008: 7(6)
- Patra JK, Dhal NK, Thatoi HN. In vitro bioactivity and phytochemical screening of *Suaeda maritima* (Dumort): A mangrove associate from Bhitarkanika, India. Asian Pacific Journal of Tropical Medicine. 2011 Sep 1; 4(9):727-34. https://doi.org/10.1016/S1995-7645 (11)60182-X.
- Ayaz, FA. Kalioglu, A., Turgut, R... Water stress effects on the contents of low molecular weight carbohydrates and phenolic acid in *Ctenanthe setose* (RoSc.) Eichler.can. J. plant Sci. 2000, 80, 373-378. https://doi.org/10.4141/P99-005.
- Ali RM, Abbas HM. Response of salt stressed barley seedlings to phenylurea. Plant Soil and Environment. 2003 Apr 1; 49(4):158-62
- Sharma P, Jha AB, Dubey RS, Pessarakli M. Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. Journal of botany. 2012; 2012.
- Agati G, Matteini P, Goti A, Tattini M. Chloroplast-located flavonoids can scavenge singlet oxygen. New Phytologist. 2007 Apr; 174(1):77-89. https://doi.org/10.1111/j.1469-8137.2007.01986.x.
- Potterat O. Antioxidants and free radical scavengers of natural origin. Current organic chemistry. 1997 Nov 1; 1(4):415-40.
- 38. Bohnert HJ, Jensen RG. Strategies for engineering water-stress tolerance in plants. Trends in Biotechnology. 1996 Mar 1; 14(3):89-97. https://doi.org/10.1016/0167-7799(96)80929-2
- Sharma P, Jha AB, Dubey RS, Pessarakli M. Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. Journal of botany. 2012; 2012.
- Sun J, Li L, Liu M, Wang M, Ding M, Deng S, Lu C, Zhou X, Shen X, Zheng X, Chen S. Hydrogen peroxide and nitric oxide mediate K+/Na+ homeostasis and antioxidant defense in NaClstressed callus cells of two contrasting poplars. Plant Cell, Tissue and Organ Culture (PCTOC). 2010 Nov 1; 103(2):205-15. https://doi.org/10.1007/s11240-010-9768-7
- Levine A, Tenhaken R, Dixon R, Lamb C. H<sub>2</sub>O<sub>2</sub> from the oxidative burst orchestrates the plant hypersensitive disease resistance response. Cell. 1994 Nov 18; 79(4):583-93. https://doi.org/10.1016/0092-8674 (94)90544-4.
- Gomez JM, Hernandez JA, Jimenez A, Del Rio LA, Sevilla F. Differential response of antioxidative enzymes of chloroplasts and mitochondria to long-term NaCl stress of pea plants. Free Radical Research. 1999 Jan 1; 31(sup1):11-8. https://doi.org/10.1080/10715769900301261
- Taji T, Seki M, Satou M, Sakurai T, Kobayashi M, Ishiyama K, Narusaka Y, Narusaka M, Zhu JK, Shinozaki K. Comparative genomics in salt tolerance between Arabidopsis and Arabidopsisrelated halophyte salt cress using Arabidopsis microarray. Plant physiology. 2004 Jul 1; 135(3):1697-709. www.plantphysiol.org/cgi/doi/10.1104/pp.104.039909.
- Panda M, Murthy TV, Samal RN, Lele N, Patnaik AK, Chand PK. Diversity of true and mangrove associates of Bhitarkanika national

- park (Odisha), India. International Journal of Advanced Research. 2017; 5(1):1784-9.
- 45. Swaminathan MS. Mangrove Plants of Tamil Nadu.
- Arunprasath A, Gomathinayagam M. Distribution and Composition of True Mangroves Species in three major Coastal Regions of Tamilnadu, India. International Journal. 2014; 2(10):241-7.
- 47. Parida AK, Das AB, Sanada Y, Mohanty P. Effects of salinity on biochemical components of the mangrove, *Aegiceras corniculatum*. Aquatic botany. 2004 Oct 1; 80(2):77-87. https://doi.org/10.1016/j.aquabot.2004.07.005.
- Saravanavel R, Ranganathan R, Anantharaman P. Effect of sodium chloride on photosynthetic pigments and photosynthetic characteristics of *Avicennia officinalis* seedlings. Recent Research in Science and Technology. 2011 Mar 28.
- Palliyath S, Puthur JT. The modulation of various physiochemical changes in *Bruguiera cylindrica* (L.) Blume affected by high concentrations of NaCl. Acta Physiologiae Plantarum. 2018 Sep 1; 40(9):160. https://doi.org/10.1007/s11738-018-2735-3
- 50. Parida A, Das AB, Das P. NaCl stress causes changes in photosynthetic pigments, proteins, and other metabolic components in the leaves of a true mangrove, *Bruguiera* parviflora, in hydroponic cultures. Journal of Plant Biology. 2002 Mar 1; 45(1):28-36.
- Elayaraj B, Selvaraju M, Dhanam S. Physiological and biochemical responses of *Ceriops roxburghiana* Arn. Seedling under salt stress conditions.
- 52. Mwangi Theuri M, Kinyamario JI, Van Speybroeck D. Photosynthesis and related physiological processes in two mangrove species, *Rhizophora mucronata* and *Ceriops tagal*, at Gazi Bay, Kenya. African Journal of Ecology. 1999 Jun; 37(2):180-93.
- 53. Ru QM, Xiao Q, Lin P, Pei ZM, Zheng HL. Short-and long-term effects of NaCl on physiological and biochemical characteristics in leaves of a true mangrove, *Kandelia candel*. Russian Journal of Plant Physiology. 2009 May 1; 56(3):363-9. https://doi.org/10.1134/S1021443709030091.
- Yan L, Guizhu C. Physiological adaptability of three mangrove species to salt stress. Acta Ecologica Sinica. 2007 Jun 1; 27(6):2208-14. https://doi.org/10.1016/S1872-2032(07)60052-3
- Iyengar ER, Reddy MP. Photosynthesis in highly salttolerant plants. Handbook of photosynthesis. M. Pessaraki.
- Telave AB. Ecophysiological studies on Sonneratia L. from the coast of Maharashtra, India. http://nopr.niscair.res.in/jspui/handle/123456789/34899
- Prado FE, Boero C, Gallardo MR, González JA. Effect of NaCl on growth germination and soluble sugars content in *Chenopodium* quinoa Willd. Seeds.
- Gadallah MA. Effects of proline and glycinebetaine on Vicia faba responses to salt stress. Biologia plantarum. 1999 Sep 1; 42(2):249-57. https://doi.org/10.1023/A:1002164719609
- Sivasankaramoorthy S, Balasubramanian T, Amuthavalli P, Sivaraman P. Studies on the Effect of Salt Stress in *Rhizophora* apiculata Bl. Journal of Phytology. 2011 Jan 11.
- Hassanein AM. Alterations in protein and esterase patterns of peanut in response to salinity stress. Biologia plantarum. 1999 Sep 1; 42(2):241-8. https://doi.org/10.1023/A:1002112702771.
- Sugihara K, Hanagata N, Dubinsky Z, Baba S, Karube I. Molecular characterization of cDNA encoding oxygen evolving enhancer protein 1 increased by salt treatment in the mangrove *Bruguiera* gymnorrhiza. Plant and cell physiology. 2000 Nov 15; 41(11):1279-85. https://doi.org/10.1093/pcp/pcd061.
- 62. Al Maqtari Maher A. Screening of salt-stress, antioxidant enzyme, and antimicrobial activity of leave extracts of mangroves *Avicennia marina* L. from Hodaidah, Yemen. Journal of Stress Physiology & Biochemistry. 2014; 10(2).
- Pareek A, Singla SL, Grover A. Salt responsive proteins/genes in crop plants.
- 64. Alamgir AN, Ali MY. Effect of salinity on leaf pigments, sugar and protein concentrations and chloroplast ATPase activity of rice (*Oryza sativa* L.). Bangladesh Journal of Botany. 1999 Dec 1; 28(2):145-9.
- Khan F, Adnan MY, Aziz I. Metabolic implications of salt induced osmolyte accumulation in *Avicennia marina*. Pak. J. Bot. 2016 Feb 1; 48(1):29-36.

- Alia, Saradhi PP. Suppression of mitochondrial electron transport activity is the prime cause behind stress induced proline accumulation. Biochem Biophys Res Commun. 1993. 403:54-58. DOI: 10.15666/aeer/1304\_9931013.
- 67. Chen J, Xiao Q, Wang C, Wang WH, Wu FH, He BY, Zhu Z, Ru QM, Zhang LL, Zheng HL. Nitric oxide alleviates oxidative stress caused by salt in leaves of a mangrove species, *Aegiceras corniculatum*. Aquatic botany. 2014 Jul 1; 117:41-7. https://doi.org/10.1016/j.aquabot.2014.04.004.
- 68. Karim MA, Islam MA, Islam MM, Rahman MS, Sultana S, Biswas S, Hosen MJ, Mozumder K, Rahman MM, Hasan MN. Evaluation of antioxidant, anti-hemolytic, cytotoxic effects and anti-bacterial activity of selected mangrove plants (*Bruguiera gymnorrhiza* and *Heritiera littoralis*) in Bangladesh. Clinical Phytoscience. 2020 Dec; 6(1):1-2. https://doi.org/10.1186/s40816-020-0152-9
- Maharana, P. K., & Basak, U. C. Studies of Non-Enzymatic and Growth Changes in Two Vegetatively Propagated Mangrove Species i.e. Excoecaria agallocha and Cerbera manghas at NaCl Stress during Hardening. 2020
- Wang L, Pan D, Lv X, Cheng CL, Li J, Liang W, Xing J, Chen W. A multilevel investigation to discover why *Kandelia candel* thrives in high salinity. Plant, cell & environment. 2016 Nov; 39(11):2486-97. https://doi.org/10.1111/pce.12804
- Agastian P, Kingsley SJ, Vivekanandan M. Effect of salinity on photosynthesis and biochemical characteristics in mulberry genotypes. Photosynthetica. 2000 Nov 1; 38(2):287-90. https://doi.org/10.1023/A:1007266932623
- Jithesh MN, Prashanth SR, Sivaprakash KR, Parida A. Monitoring expression profiles of antioxidant genes to salinity, iron, oxidative, light and hyperosmotic stresses in the highly salt tolerant grey mangrove, Avicennia marina (Forsk.) Vierh. By mRNA analysis. Plant cell reports. 2006 Aug 1; 25(8):865-76. https://doi.org/10.1007/s00299-006-0127-4
- 73. Li N, Li C, Chen S, Chang Y, Zhang Y, Wang R, Shi Y, Zheng X, Fritz E, Hüttermann A. Abscisic acid, calmodulin response to short term and long term salinity and the relevance to NaCl-induced antioxidant defense in two mangrove species. The Open Forest Science Journal. 2009 Nov 4; 2(1). DOI: 10.2174/1874398600902010048
- Kavitha K, George S, Venkataraman G, Parida A. A salt-inducible chloroplastic monodehydroascorbate reductase from halophyte *Avicennia marina* confers salt stress tolerance on transgenic plants. Biochimie. 2010 Oct 1; 92(10):1321-9. https://doi.org/10.1016/j.biochi.2010.06.009
- Lu Y, Li N, Sun J, Hou P, Jing X, Zhu H, Deng S, Han Y, Huang X, Ma X, Zhao N. Exogenous hydrogen peroxide, nitric oxide and calcium mediate root ion fluxes in two non-secretor mangrove species subjected to NaCl stress. Tree physiology. 2013 Jan 1; 33(1):81-95. https://doi.org/10.1093/treephys/tps119.
- Lang T, Sun H, Li N, Lu Y, Shen Z, Jing X, Xiang M, Shen X, Chen S. Multiple signaling networks of extracellular ATP, hydrogen peroxide, calcium, and nitric oxide in the mediation of root ion fluxes in secretor and non-secretor mangroves under salt stress. Aquatic botany. 2014 Oct 1; 119:33-43. https://doi.org/10.1016/j.aquabot.2014.06.009
- 77. Nirjhar Dasgupta PN, Sengupta C, Das S. Occurrence of Secondary Metabolites and Free Radical Scavenging Ability towards Better Adaptability of Some Mangrove Species in Elevated Salinity of Indian Sundarbans. Annals of Tropical Research. 2017; 39(1):12-40.
- Asaeda T, Barnuevo A. Oxidative stress as an indicator of nichewidth preference of mangrove *Rhizophora stylosa*. Forest Ecology and Management. 2019 Jan 15; 432:73-82. https://doi.org/10.1016/j.foreco.2018.09.015.
- Ksouri R, Megdiche W, Debez A, Falleh H, Grignon C, Abdelly C. Salinity effects on polyphenol content and antioxidant activities in leaves of the halophyte *Cakile maritima*. Plant Physiology and Biochemistry. 2007 Mar 1; 45(3-4):244-9. https://doi.org/10.1016/j.plaphy.2007.02.001.